



Plant sampling summary

APAL 

Australian Precision Ag Laboratory

www.apal.com.au

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**Correct sampling is imperative to ensure accurate analysis and interpretation.
Precise nutrient management can only occur with a quality representative sample.**

**Sample the correct part of the plant at the correct growth stage (see next page)
Always wear gloves to avoid contamination**

Regular tests build better profile

Things to avoid:

- Avoid spoiled, diseased, insect damaged, senescing or dead plant tissue
- Avoid unusual areas in the paddock which can include poorly drained soils, sheep camps, headlands etc. (as per soil sampling guidelines)
- Avoid areas of crops/pastures in the immediate vicinity of trees
- Don't sample plants under moisture stress and avoid waterlogging
- Avoid plants growing in dung or urine patches
- Avoid sampling soon after soil or foliar fertiliser application
- Avoid contamination from dust, soil, sprays, fertilisers and your hands (Always wear gloves to avoid contamination)
- Be consistent in obtaining your representative sample, sample plants of similar vigour, variety, size and age
- Sample early in the week; if storing samples over the weekend, dry or refrigerate
- Avoid plastic bags; use APAL's paper bags. If you require bags please contact us

Taking the sample

1. Review your required tests from APAL's service guide. If guidance is required, consult APAL or your local adviser/agronomist
2. Determine an appropriate sampling plan
3. Ensuring a representative sample is obtained
4. Avoid unusual areas listed above
5. When trouble shooting take samples from good and poor growth areas to compare
6. Ensure you are wearing gloves to avoid contamination. Avoid touching fence posts or anything that can cause contamination between samples. Use stainless steel cutting implements
7. Ensure the correct part of the plant is sampled. See back page for summary or www.apal.com.au
8. Collect the desired amount of leaves or petioles (generally aim to 1/2 to 2/3 fill APAL's plant tissue bags). Make sure sample numbers are representative of the intended sampling area

Sending the sample to the lab

1. Ensure you have an adequate sample
2. Ensure you have submitted the correct information into APAL Online or Farm2Lab

Deliver to:

**Australian Precision Ag Laboratory
PO Box 155 Welland
South Australia 5007**

For any further information please contact APAL on 08 8332 0199 or info@apal.com.au

Orchards / Tree Crops / Vegetable Crops:

Select the most recent fully developed leaf
Usually 3rd or 4th from the growing tip.



Grape Vines:

(Please separate leaf blades from petioles before sending)

Leaf blade - sample 20-30 most recently mature leaf blades from each representative area.

(remove petiole immediately after sampling blade).

Leaf petiole - at flowering collect 40-50 petioles from leaves opposite the basal bunch from each representative area
(remove blades from petioles before sending).



Olives:

Select fully expanded mid-shoot leaves from new season growth.
Collect 4 leaves per tree from 25 trees of a single cultivar.

Potatoes:

Select the petiole (or petiole & blade) of the 5th leaf from the growing terminal.
*Always include growth stage information - ie. tuber size - length of the longest tuber.

Cereals:

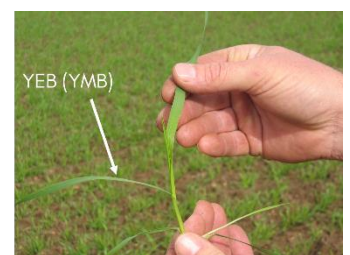
Whole tops - ensure roots and soil are removed to avoid contamination.

Take the correct part of the plant - the youngest fully emerged leaf blade (YEB or youngest mature blade - YMB) with the ligule visible.

Do not take the seed head if it has emerged.

Lupins/beans/canola:

Collect 100 of the youngest mature leaves.



Deciduous fruits:

In mid-summer collect 50 leaves from a mid-shoot position.

Cotton: Sample prior to or at first bloom or when the first squares appear.
Youngest fully matured leaves on the main stem. ~ 50-70 leaves



More crop information is available in APAL's plant sampling instructions at www.apal.com.au